Journal of Hard Tissue Biology 31[2] (2022) 63-70 2022 The Hard Tissue Biology Network Association Printed in Japan, All rights reserved. CODEN-JHTBFF, ISSN 1341-7649

The Effect of Different Surgical Instruments for Bone Regeneration under the Surgery of Bone Defect on Rat Calvaria

Takashi Furumori¹⁾, Mamoru Ueda^{2,3)}, Yoshitomo Honda⁴⁾, Yoshiya Hashimoto³⁾, Tadasuke Tanioka¹⁾, Kaoru Kusano¹⁾ and Shunsuke Baba¹⁾

¹⁾ Department of Oral Implantology, Osaka Dental University, Osaka, Japan

²⁾ Second Department of Oral and Maxillofacial Surgery, Osaka Dental University, Osaka, Japan

³⁾ Department of Biomaterials, Osaka Dental University, Osaka, Japan

⁴⁾ Department of Oral Anatomy, Osaka Dental University, Osaka, Japan

(Accepted for publication, January 21, 2022)

Abstract: In this study, we compared the healing process of bone defects treated with a trephine bur with those treated with an ultrasonic knife using a critical-sized bone defect model on rat calvaria. Nine-millimeter critical-size bone defects were prepared using both instruments in the calvaria of adult Sprague-Dawley rats. One and four weeks after the osteotomy, we performed a histomorphometric analysis to evaluate bone regeneration around the cutting surface. Quantitative micro-computed tomography analyses of the bone volume in both groups suggested that ultrasonic knife surgery resulted in superior bone formation compared to that in trephine bur surgery. Furthermore, at the cutting surface, the ultrasonic knife treatment retained the alkaline phosphatase activity and new bone formation, which was identified using calcein staining, even one week after surgery. Considering the speed and volume of bone regeneration, the ultrasonic knife is likely to be the preferred over the trephine bur to perform osteotomies in implant surgery.

Key words: Bone regeneration, Surgical instruments, Calvaria, Ultrasonic knife, Trephine bur

Introduction

In dental implantology, osteoblast attachment in the early phase is necessary, as it significantly influences implant fixation and is associated with implant survival¹⁾. The placement of a dental implant in the bone activates a sequence of molecular and cellular events that lead to the apposition of newly formed bone directly onto the titanium surface²⁾. The surface structure of the implanted material and the condition of the surface of the implant bed both affect osteointegration³⁾. Dental implant failures that occur clinically for unknown reasons could be attributed to undiagnosed damage to the bone surface of the implant bed⁴⁾. Conventional rotary burs are frequently used for preparing implant drilled holes. Major problems encountered during bone drilling are thermal necrosis, bur deformation, and microcrack generation on the inner surface of the drilled holes, which can detrimentally affect the subsequent healing process⁵⁾.

Piezosurgery using ultrasonic vibrations has been used for osteotomy since the 1950s as an alternative technique for surgery using a rotary bur^{6,7)}. Ultrasonic instruments were used for implant surgery because they reduce the incidence of bone burns, and because these instruments could selectively cut the mineralized tissue⁴⁾. Surgery using ultrasonic instruments is known to reduce soft tissue damage because of the selective cutting that can be performed on mineralized tissue when the instruments are used at 25–30 kHz⁸⁻¹⁰⁾. Clinical studies have shown that ultrasonic techniques can provide a clear view of the surgical field, leading to reduced damage to essential anatomic structures such as the nerves or

blood vessels^{11,12}, reduced damage to the tissue wound leads to faster healing¹³. In contrast, the conventional method that uses rotary drills in preparing implant sites causes thermal damage to the tissue in the implant bed, which may lead to reduced fixation and worse osteointegration around the dental implant¹⁴. However, the differences in the bone regeneration process after osteotomy surgery using an ultrasonic knife or conventional rotary bur at the same surgical time remain controversial.

This study aimed to compare the effects of an ultrasonic knife and a trephine bur on bone regeneration used for similar durations on a rat critical-size defect model.

Materials and Methods

Rat calvarial-defect model

Sixteen male Sprague-Dawley rats (8 weeks old, 250–270 g, SHIMI-ZU Laboratory Supplies Co, Ltd., Kyoto, Japan) were used for the animal studies. The experimental protocol was approved by the Animal Care and Use Committee of Osaka Dental University (approval number: 17-03008). The rats were divided into the following groups: (i) the trephine bur group, further divided into the 1- and 4-week groups (n = 4/group), and (ii) the ultrasonic knife group, further divided into the 1- and 4-week groups (n = 4/group) (Fig. 1). General anesthesia was induced by a combination of butorphanol, midazolam, and medetomidine by intraperitoneal injection, followed by local anesthesia using lidocaine by subcutaneous injection in the surgical area. Hair clipping was performed outside the operation area using an electric clipper and hair removal cream. The skin was prepared using iodine, which was subsequently wiped off using 70% ethanol, which was sprayed on the surgical site. A 9-mm defect was created using a trephine bur (Implatex Co, Ltd.,

Original

Correspondence to: Dr. Takashi Furumori, Department of Oral Implantology, Osaka Dental University 1-5-17, Otemae, Chuo-ku, Osaka-shi, Osaka 540-0008, Japan; Tel: +81-6-6910-1520; Fax: +81-6-6910-1048; E-mail: furumoridc@gmail.com

J.Hard Tissue Biology Vol. 31(2): 63-70, 2022



Figure 1. Surgical preparation of the defects. (a, b) U-shaped incisions on skin and periostin made to explore the calvarial bones; (c) bony defect is marked on the centre of the calvaria using a 9-mm trephine bur; (d) defect prepared using an ultrasonic knife with a scale-type cutting edge; (e) mucosal elevator was used to remove the bone fragments; (f) bony defect prepared using the trephine bur; (g) defect prepared using an ultrasonic knife; (h) periosteum was repositioned and sutured; (i) skin is closed.

Tokyo, Japan) or an ultrasonic knife (Sonic Surgeon 310 L, Dong IL Technology, Gyeonggi-do, Korea) with water injected in the center of the calvaria as a coolant. A mucosal elevator was used to remove the bone fragments. The periosteum was subsequently repositioned using a 5-0 suture, and the skin was repositioned using a 4-0 suture. The animals received a postoperative antibiotic regimen of gentamicin (GENTACIN[®], MSD K.K., Tokyo, Japan) for two days (2 mg/kg/day). Further, four rats in the trephine bur group or ultrasonic knife group were injected with calcein after surgery (2 mg/kg; Wako Pure Chemical Industries Co., Osaka, Japan), which was continued for three days; these rats were euthanized at seven days after surgery. The other rats were euthanized using an overdose of pentobarbital sodium (2 ml/kg, Somnopentyl[®], Kyoritsu, Tokyo, Japan) one month after surgery. The wounds were observed without signs of infection, dehiscence, or self-inflicted trauma.

Scanning electron microscopic observation

The cut edges of the surgical site that were made using a trephine bur or ultrasonic knife were observed using a field emission-scanning electron microscope (FE-SEM; 5-kV, S-4800, Hitachi High Technologies, Tokyo, Japan).

Bone histomorphometry

The samples were fixed in 4% paraformaldehyde for 24 h. The Kawamoto method was used to obtain four-micrometer-thick non-decalcified frozen sections¹⁵⁾. The dynamic osteogenesis was studied by observing the fluorescently labeled sections from the 4-week group under an LSM700 laser-scanning microscope (Zeiss, Jena, Germany). To activate the fluorophores, lasers of different wavelengths were used, namely 488 nm (calcein, yellow) or 555 nm (Alizarin Red, red).

Histochemical staining and histological observations

Alkaline phosphatase (ALP) and tartrate-resistant acid phosphatase (TRAP) were stained for histological analysis. The TRAP and ALP staining were performed using the TRAP/ALP Kit (Wako Pure Chemical Industries Co., Osaka, Japan) to identify the osteoclasts and measure osteoblast activity. In our previous studies^{16,17}, we confirmed TRAP

staining in frozen sections. So, we used the same method. After staining, the sections were observed using a BZ-9000 digital microscope (Keyence Co., Osaka, Japan).

Statistical Analysis

Statcel3 software (OMS, Tokyo, Japan) were used for the statistical analysis. For all experiments, values are reported as the mean \pm standard deviation (SD). For comparisons between the two groups, the data were evaluated using Student's t-test. Statistical significance was set at p < 0.05.

Results

Operation time and bleeding volume

Although surgery using an ultrasonic knife requires more time to complete¹⁸⁻²⁰⁾, no significant difference was observed between the trephine bur and ultrasonic knife groups in terms of bleeding volume and operation time because of the significant deviation in the trephine bur



Figure 2. Quantification of operation time and bleeding volume of using trephine bur or ultrasonic knife. No significant difference could be found (SD; n = 8). Statistically significant difference at p < 0.05 (Student's t-test).

Trephine bur

Ultrasonic knife



Figure 3. Scanning electronic micrographic images at the margin of the defects after preparation with trephine bur or ultrasonic knife (a, b).



Figure 4. Critically sized bone defects in rat calvaria treated with the ultrasonic knife or trephine bur. Micro-computed tomography (a) and bone-mineral density (BMD) (b) images of the defects in the rat calvaria. (c) Postoperative bone volumes/tissue volumes (BV/TV). Black squares: Ultrasonic knife; Gray circle: Trephine bur. The data shown represent the mean \pm standard deviation (SD; n = 4); ** *p* < 0.01 (Student's t-test).

group (Fig. 2). The standard deviation of the ultrasonic knife group was smaller than that of the trephine bur group.

SEM

In the SEM images of the trephine burr group (Fig. 3), the cutting surface of the bone defect was rough, with a large amount of bone debris (white arrows). However, in the ultrasonic knife group, the cutting surface was smooth.

Micro-computed tomography

We examined the bone morphometric changes that occurred after surgery (Fig. 4). In the trephine bur group, micro-computed tomography images and structural parameters of rat calvaria showed that the volume of the new bone formed on the bone defects in the ultrasonic knife group was greater than that formed in the trephine bur group. The trephine bur group showed a significantly lower average bone volume than the ultrasonic surgery group at four weeks (p < 0.01).

Fluorescence imaging

Fig. 5 shows the early bone formation identified using calcein (green) at the cutting surface of defects 1 week after surgery (Fig. 5). In addition, we found that different bone formations occurred on the cutting surface of the ultrasonic knife group, and the trephine bur group showed a reduced new bone formation compared to that in the ultrasonic knife group.

ALP/TRAP staining

We further evaluated the ALP/TRAP expression using histochemistry to assess the bone turnover capacity (ALP and TRAP staining for detecting osteoblast and osteoclast activation, respectively) in each group (Fig. 6 for ALP and Fig. 7 for TRAP staining). One week after treatment, the trephine bur group samples showed weak ALP expression in the cutting surface. In contrast, the samples from the ultrasonic knife groups were significantly stained black (ALP expression), indicating that the surgery using the ultrasonic knife could help retain bone-forming ability (Fig. 6). Further, the samples in the trephine bur group



Figure 5. Fluorescence labeling analysis. Calcein (green staining: newly formed bone at 1-week post-implantation) and Alizarin red (red: contrastaining of calcified tissue) labeling of the bone tissue in the calvarial defects.

1 W

4 W



Ultrasonic knife

Trephine bar

Figure 6. Alkaline phosphatase (ALP) staining; black stains represent ALP-positive tissue. OB: Original Bone, NB: New Bone.



Figure 7. Tartrate-resistant acid phosphatase (TRAP) staining showing the presence of osteoclasts in the tissue sections.

showed significant TRAP staining (black arrow in Fig. 7) in the bone debris, suggesting that in cases of trephine bur surgery, bone resorption was promoted to remove the damaged bone tissue before the initiation of defect healing. This suggests surgery using an ultrasonic knife may help attenuate damage to the mother bone, leading to bone regeneration being initiated earlier than in cases where the trephine bur is used.

Discussion

Some studies comparing the efficacy of osteotomy using an ultrasonic knife and with that of conventional techniques using a trephine bur for bone healing have reported no differences in the postoperative bone formation^{21,22)}. However, in the present study, we demonstrated that critical-size bone defects formed with a trephine bur and an ultrasonic knife induced bone formation at different speeds. Postoperative bone formation in cases of ultrasonic surgery was significantly greater and began earlier than that in cases of surgery performed using a trephine bur in a rat model.

In our experimental rat model, the differences in bleeding or operation time between osteotomies prepared by ultrasonic knife and conventional rotary burs failed to reach statistical significance (Fig. 2). However, this may be because the surgeon using the trephine bur must have been cautious in avoiding damage to the dorsal cerebral vein. Generally, if the trephine bur comes in contact with the dorsal cerebral vein, it causes severe hemorrhage, which is difficult to control during the surgery, which increases the duration of the surgery. In addition, damage to the dorsal cerebral vein, which causes large blood clots and swelling, is likely to influence bone repair progress. However, the narrow and long ultrasonic knife provides a clearer and unobscured view of the surgical field, including delicate vein structures, which can be used to circumvent severe hemorrhage.

In clinical practice, osseointegration to the implant is affected by the surface structure of the implant body and the condition of the surface of the implant bed¹²⁾. It is believed that protecting the surface of implant bed from damage equally important²³⁾. In the present study, at the cutting surface of bone defects treated with trephine bur, we observed more bone fragments, suggesting that increased damage was caused to the bone surface compared to that in ultrasonic knife surgery. In cases where the trephine bur was used, osteoclastic absorption of the bone regeneration is likely to take some time. In contrast, a smaller amount of bone debris was observed at the cutting surface in cases of ultrasonic knife than that in trephine bur surgery, suggesting that the former tech-

nique causes reduced damage to the bone defect surface (Fig. 3). These results indicate that the early initiation of bone healing in cases of ultrasonic knife surgery can be attributed to reduced damage to the mother bone.

Vajgel et.al. reported a systematic review that showed that when using the conventional rotary drill, the mean expected new bone formation in 9.0-mm-diameter calvaria defects in the rat model was 11.18% of that at one month after surgery²⁴⁾. The mean bone formation can range from 4.93%-30%, depending on the surgeon. In the present study, the defect prepared using ultrasonic surgery was 35.1% one month after surgery. Additionally, we believe that the ultrasonic knife reduced the risk of damage to the soft tissues when compared with that of the round bur²⁵⁾. Considering these benefits, the ultrasonic knife may be a better candidate for osteotomy, to help induce earlier bone formation. However, in our study, one surgeon performed all the osteotomies. The deviation of bone regeneration was 19.7%-53.1% when an ultrasonic knife was used without any bone grafts or bone substitutes. It is believed that the skill of a surgeon significantly influences the quality of surgery. Further detailed investigations involving more surgeons are required to conclusively prove the superiority or inferiority of these instruments.

The experimental model with a critical-sized bone defect in calvaria is a well-used model in mice, rats, rabbits, and canines to detect the effect of certain devices²⁶⁻²⁹⁾ and elucidate the bone-forming ability of autogenous bone grafts or of various biomaterials³⁰⁻³³⁾. In this study, bone defects formed using the trephine bur showed a reduced deviation than those formed using an ultrasonic knife (Fig. 4), suggesting that a trephine bur is likely to be a suited for the preparation of a severe and consistent bone defect model in rat calvaria when comparing various materials. Our results indicate that careful attention should be paid to the surgical devices before comparing the results of bone formation in different studies, even for bone defects of the same size; the use of different surgical devices may alter the basement level of bone formation.

In conclusion, this study shows that bone regeneration in rat calvaria can be enhanced when using an ultrasonic knife in preparing osteotomy defects compared to using trephine bur under similar surgical times. The increased bone formation in cases of ultrasonic knife surgery may be due to the reduced damage to the mother bone, possibly due to the sustained bone-forming activity of osteoblasts. In addition to other advantages of ultrasonic knives, such as minor damage to soft tissues, the results of our study suggest that ultrasonic knife surgery may be preferable to conventional osteotomy surgery in clinical settings.

Conflicts of Interest

The authors have declared that no COI exist.

References

- Meyer U, Joos U, Mythili J, Stamm T, Hohoff A, Fillies T, Stratmann U and Wiesmann H.P. Ultrastructural characterization of the implant/bone interface of immediately loaded dental implants. Biomaterials 25: 1959-1967, 2004
- Meyer U, Wiesmann, H.P, Fillies T and Joos U. Early tissue reaction at the interface of immediately loaded dental implants. Int J Oral Maxillofac Implants 18: 489-499, 2003
- Delgado-Ruiz RA, Ortega EV, Romanos GE, Gerhke S, Newen I and Calvo-Guirado JL. Slow drilling speeds for single-drill implant bed preparation in vitro study. Clin Oral Investig 22: 349-359, 2018
- Scarano A. Traditional postextractive implant site preparation compared with pre-extractive interradicular implant bed preparation in the mandibular molar region, using an ultrasonic device: A randomized pilot study. Int J Oral Maxillofac Implants 32: 655-660, 2017
- Fugito JK, Cortes AR, Ricardo CD and Yoshimoto M. Comparative study on the cutting effectiveness and heat generation of rotary instruments versus piezoelectric surgery tips using scanning electron microscopy and thermal analysis. Int J Oral Maxillofac Implants 33: 345-350, 2018
- Horton JE, Tarpley, TMJr and Wood LD. The healing of surgical defects in alveolar bone produced with ultrasonic instrumentation, chisel, and rotary bur. Oral Surg Oral Med Oral Pathol 39: 536-546, 1975
- Stubinger S, Stricker A and Berg BI. Piezosurgery in implant dentistry. Clin Cosmet Investig Dent 7: 115-124, 2015
- Labanca M, Azzola F, Vinci R and Rodella LF. Piezoelectric surgery: twenty years of use. Br J Oral Maxillofac Surg 46: 265-269, 2008
- Schlee M, Steigmann M, Bratu E and Garg AK. Piezosurgery: basics and possibilities. Implant Dent 15: 334-340, 2006
- Robiony M, Polini F, Costa F, Vercellotti T and Politi M. Piezoelectric bone cutting in multipiece maxillary osteotomies. J Oral Maxillofac Surg 62: 759-761, 2004
- Scarano A, Carinici F, Lorusso F, Festa F, Bevilacqua L, Oliveria PS and Maglione M. Ultrasonic vs drill implant site preparation: post-operative pain measurement through VAS, swelling and crestal bone remodeling: A randomized clinical study. Materials 11(12): 2516, 2018
- Scarano A, Lezzi G, Perrotti V, Tetè S, Staiti G, Mortellaro C and Cappucci C. Ultrasonic versus drills implant site preparation: a histologic analysis in bovine ribs. J Craniofac Surg 25: 814-817, 2014
- Moslemi N, Shahnaz A, Masoumi S, Torabi S and Akbari S. Laser-assisted osteotomy for implant site preparation: A literature review. Implant Dent 26: 129-136, 2017
- 14. Vigano P, Botticelli D, Salata LA, Schweikert MT, Velez JU and Lang NP. Healing at implant sites prepared conventionally or by means of Sonosurgery[®]. An experimental study in dogs. Clin Oral Implants Res 26: 377-382, 2015
- Kawamoto T. Use of anew adhesive filn for the preparation of multi-purpose fresh-frozen sections from hard tissues, whole-animals, insects and plants. Arch Histol Cytol 66(2): 123-143, 2003
- Hieda A, Uemura N, Hashimoto Y, Toda I and Baba S. *In vivo* bioactivity of porous polyetheretherketone with a foamed surface. Dent Mater J 36(2): 222-229, 2017

- Li P, Hashimoto Y, Honda Y, Arima Y and Matsumoto N. The effect of interferon-γ and zoledronate treatment on alpha- tricalcium phosphate/collagen sponge- mediated bone- tissue engineering. Int J Mol Sci 16: 25678-25690, 2015
- Iacoangeli M, Neri P, Balercia P, Lupi E, Di Rienzo A, Nocchi N, Alvaro L and Scerrati M. Piezosurgery for osteotomies in orbital surgery: Our experience and review of the literature. Int J Surg Case Rep 4: 188-191, 2013
- Sortino F, Pedulla E and Masoli V. The piezoelectric and rotatory osteotomy technique in impacted third molar surgery: Comparison of postoperative recovery. J Oral Maxillofac Surg 66: 2444-2448, 2008
- 20. Bartuli FN, Luciani F, Caddeo F, Chiara LDE, Di Dio M, Piva P, Ottria L and Arcuri C. Piezosurgery vs high speed rotary handpiece: A comparison between the two techniques in the impacted third molar surgery. Oral Implantol (Rome) 6(1): 5-10, 2013
- Esteves JC, Marcantonio EJ, Faloni APS, Rocha FRG, Marcantonio RA, Wilk K and Intini G. Dynamics of bone healing after osteotomy with piezosurgery or conventional drilling – histomorphometrical, immunohistochemical, and molecular analysis. J Transl Med 11: 221, 2013
- 22. Ma L, Stubinger S, Liu XL, Schneider UA and Lang NP. Healing of osteotomy sites applying either piezosurgery or two conventional saw blades: a pilot study in rabbits. Int Orthop 37: 1597-1603, 2013
- Shibata Y, Tanimoto Y, Maruyama N and Nagakura M. A review of improved fixation methods for dental implants. Part II: biomechanical integrity at bone-implant interface. J Prosthodont Res 59: 84-95, 2015
- Vajgel A, Mardas N, Farias BC, Petrie A, Cimões R and Donos N. A systematic review on the critical size defect model. Clin Oral Implants Res 25: 879-893, 2014
- 25. Otake Y, Nakamura M, Henmi A, Takahashi T and Ssano Y. Experimental comparison of the performance of cutting bone and soft tissue between piezosurgery and conventional rotary. Sci Rep 8: 17154, 2018
- Kaida K, Honda Y, Hashimoto Y, Tanaka M and Baba S. Application of green tea catechin for inducing the osteogenic differentiation of human dedifferentiated fat cells *in vitro*. Int J Mol Sci 16: 27988-28000, 2015
- 27. Honda Y, Takeda Y, Li P, Huang A, Sasayama S, Hara E, Uemura N, Ueda M, Hashimoto M, Arita K, Matsumoto N, Hashimoto Y, Baba S andTanaka T. Epigallocatechin gallate-modified gelatin sponges treated by vacuum heating as a novel scaffold for bone tissue engineering. Molecules 23: 876, 2018
- Aaboe M, Pinholt EM, Schou S and Hjorting-Hansen E. Incomplete bone regeneration of rabbit calvarial defects using different membranes. Clin Oral Impl Res 9: 313-320, 1998
- 29. Mahesh HM, Sergei AK, Brian S, Ravi KN, Robert OR, Yixian Q and Pamela GR. Canine cranial reconstruction using autologous bone marrow stromal cells. Am J Pathol 168: 542-550, 2006
- Nasrin E, Parsa D, Nasrin T, Mehrnoosh R and Niloufar D. Histologic evaluation of the bone regeneration capacities of bio-oss and mineross X in rabbit calvarial defects. Int J Periodontics Restorative Dent 39: 6, 2019
- Nakano K, Kubo H, Nakajima M, Honda Y and Hshimoto Y. Bone regeneration using rat-derived dedifferentiated fat cells combined with activated platelet-rich plasma. Materials 13: 5097, 2020
- 32. Hara E, Honda Y, Suzuki O, Tanaka T and Matsumoto N. Epigallocatechin gallate-modified gelatins with different compositions alter

the quality of regenerated bones. Int J Mol Sci 19: 3232, 2018

33. Takeda T, Honda Y, Kakinoki S, Yamaoka T and Baba S. Surface modification of porous alpha-tricalcium phosphate granules with

heparin enhanced their early osteogenic capability in a rat calvarial defect model. Dent Mater J 37(4): 575-581, 2018

J.Hard Tissue Biology Vol. 31(2): 63-70, 2022